

Detection of LNA-containing antisense oligonucleotides in rat liver and kidney by immunogold transmission electron microscopy

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1. Introduction

Locked Nucleic Acids (LNA) are chemically modified RNA-like nucleotides with the potential to be used in therapeutic antisense oligonucleotides (ASO) to prevent expression of “disease-related” protein products.

ASO mode of action

LNA structure

Gapmer LNA design

Despite their proven therapeutic potential, some LNA-containing ASO have shown **toxic** effects mainly associated with off-target effects and with their accumulation in liver and kidney.

General aim: To investigate pathogenic mechanisms of toxicity of selected tool LNA ASO with different safety profiles, comparing unconjugated (naked) LNA ASO or conjugated with *N*-acetyl galactosamine (GalNAc) for targeted delivery to hepatocytes.

Focus of this poster: To establish an immunogold methodology to investigate the accumulation of LNA in liver and kidney at subcellular level.

2. Materials and Methods

Study design

Safety profile (toxicity)	Tool compounds		Target (sequence)	Dosing (weekly)
	Naked	GalNAc		
Control	Vehicle			• SC administration
Non toxic	ASO1	Not tested	None, scramble (sequence 1)	• Sacrificed after 1, 3 or 6 doses
Moderately toxic	ASO2	ASO3	Human PCSK9 (sequence 2)	• Sacrificed 3 days after last dose
Highly toxic	ASO4	ASO5	Human PCSK9 (sequence 3)	• Naked LNAs: 40 mg/kg • GalNAc LNAs: 20 mg/kg

Species/Sex: Wistar rat (male). Special sampling of liver & kidney

Immunogold TEM workflow

Immunogold labelling (main steps):

Workflow steps:

Protocol details:

3. Results (I): KIDNEY

Abbreviations: BB (brush border), EGPV (electron-dense gold-positive vesicles = LNA ASO), G (glomerulus), M (mitochondrion), N (nucleus), PTEC/DTEC (proximal/distal tubular epithelial cells).

A (ASO4, 3 doses)

B (ASO2, 1 dose)

C (Vehicle, 1 dose)

D (ASO4, 1 dose)

E (ASO3, 1 dose)

F (ASO4, 1 dose)

G (ASO2, 1 dose)

H (ASO5, 3 doses)

Semithin section stained with toluidine blue, showing the LNA granules in PTEC (dark blue, inset), but not in DTEC (arrowhead) or glomeruli (A). LNA-containing ASO are observed as **electron-dense gold-positive vesicles** (EGPV) with **irregular shape**, consistent with endosomes or secondary lysosomes (B, E-H) [1,2].

Electron-dense **protein droplets** (gold negative) compatible with secondary lysosomes loaded with *alpha 2u-globulin*, are observed in both control (C) and dosed rats (D). Protein droplets can get larger and polyangular-shaped, and crystalloids can form with characteristic strial pattern (D inset) [3].

Some organelles undergoing degradation (mitochondria (E, arrowheads), ribosomes and RER (F, arrow)) were observed within the same membranes containing the LNA vesicles, suggesting (at least partially) common pathways with **autophagy**.

Differences in the density of gold particles were observed within the same sample (G, top vs bottom vesicles), and among compounds (G vs H), probably due to different stages of LNA degradation, and to differences in the antibody binding affinity to the compounds, respectively. ASO2&3 showed the strongest gold reactivity (B, E, G).

EGPV are observed surrounded by a moderately electron-dense material (grey) suggestive of lysosomal content (⬢), and/or by electron-lucent material (white) suggestive of osmotic hydration (⬢).

4. Results (II): LIVER

Abbreviations: EC (sinusoid endothelial cell), EGPV (electron-dense gold-positive vesicles = LNA ASO), H (hepatocyte), IC (Ito cell), KC (Kupffer cell) M (mitochondrion), N (nucleus).

A (Vehicle, 1 dose)

B (Vehicle, 3 doses)

C (ASO2, 3 doses)

D (ASO3, 3 doses)

E (ASO5, 1 dose)

F (ASO5, 3 doses)

Figures A&B: No electron-dense gold-positive vesicles (EGPV) were observed in any cell type in control rats, confirming the specificity of the labelling.

Figures C&D: EGPV within **Kupffer cells** in rats dosed with both **naked** (C) and **GalNAc**-conjugated LNAs (D). LNAs in Kupffer cells show a more **round morphology**, reflecting phagocytic uptake (phagosomes), unlike LNAs in the kidney, where more irregular shapes are observed, most likely as a result of different uptake mechanisms (endocytic uptake, endosomes) [1,2].

Figures E&F: EGPV within **hepatocytes** in rats dosed with **GalNAc**-conjugated LNAs. Figure F shows features of highly degenerated hepatocytes, due to dosing with highly toxic GalNAc-conjugated LNAs.

5. Conclusions: Different morphologies of LNA vesicles may reflect different uptake mechanisms (endocytosis vs phagocytosis). Different stages of LNA degradation/metabolism and differences in the antibody binding affinity to the different compounds may explain the differences in the density of gold particles. EGPV were detected in kidney PTEC, as well as in Kupffer cells (naked and GalNAc-conjugated compounds) and hepatocytes (GalNAc-conjugated LNAs). This methodology provides new insights on the accumulation of LNAs at ultrastructural level.

6. References:

[1] Geary RS et al., 2015. Pharmacokinetics, biodistribution and cell uptake of antisense oligonucleotides. *Advanced drug delivery reviews*.

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[3] Read NG, 1991. The role of lysosomes in hyaline droplet nephropathy induced by a variety of pharmacological agents in the male rat. *Histochem. J.* 23, 436-443.